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Effects of coumarin substituents on the photophysical properties of newly synthesised phthalocyanine derivatives

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In this study, synthesis of new ligands, 8-hydroxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin and 8-hexyloxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin, and their phthalocyanines, 2,9,16,23-tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]metal-free and metallophthalocyanines {M[Pc(OBzCou)_4] (M = 2H, Zn(II), Co(II); Bz: benzene; Cou: coumarin)} were synthesised. The novel chromogenic compounds were characterised by elemental analysis: ¹H NMR, ¹³C NMR, MALDI-TOF, IR and UV–vis spectral data. The effect of coumarin substituents on the photophysical properties of metal-free (H₂Pc) and zinc phthalocyanines (ZnPc) derivatives has been examined. Spectrophotometric measurements revealed that coumarinsubstituted ZnPc derivatives were in the unaggregated form, whereas those of H₂Pc species were in aggregated form. It means that substitution of coumarin derivative prevents the cluster formation in the presence of zinc ion in the centre of Pc.

Keywords: phthalocyanines; fluorescence; coumarin derivatives; aggregation; cobalt; zinc

Introduction

Phthalocyanines exhibit excellent chemical and photochemical properties, but poor fluorescent properties (1-3). An approach to the problem in the spectrofluorometry that measures emission and excitation intensities of a fluorescent molecule was the introduction of metalchelating groups into fluorescent dyes with the aim of forming derivatives that would undergo changes in the fluorescence intensity and/or wavelength upon formation of metal complexes (3-5). Detection of various metal ions is therefore crucial in biological as well as electronic areas and there are numerous detection techniques in use today (6). The benefits of phthalocyanines (Pcs) in terms of technological applications are beyond question, such as sensing elements in chemical sensors, photoconducting agents in photocopying machines and carrier generation materials in near-IR (7-10). Although several thousands of phthalocyanine complexes have been synthesised and tested so far for technological applications since the discovery of phthalocyanine, many important properties have, however, been extensively studied for their low solubility in most organic solvents and their aggregation (11). It is well known that their physicochemical properties can be fine-tuned by changing the metal and/or nature of the substituents on the periphery of the Pcs(12, 13). On the other hand, the family of the functional phthalocyanines has been an interesting target for the chemists for the development of further chemical reactions on the phthalocyanine complexes (14, 15). Therefore, in this paper, we have in particular combined functional coumarins (2H-1-benzopyran-2-one) with phthalocyanines as a single compound via synthetic methodology to form soluble phthalocyanine-bearing coumarin derivative as a fluorescence probe. The experimental results demonstrate that introduction of coumarin derivatives into the peripheral ring of phthalocyanines enhances the fluorescence property of phthalocyanines. Phthalocyanines coupled with the coumarin moiety may exhibit biological activities, and after the cleavage of the lactone ring of the peripheral 2H-1-benzopyran-2-one (coumarin) in the phthalocyanine, the solubility can also be increased (*16*).

In this study, metal-free and metallophthalocyanine derivatives, {M[Pc(OBzCou)₄] (M = 2H, Zn(II), Co(II))}, containing 8-hexyloxy-3-(4-oxyphenyl)coumarin and the cleavage of the lactone ring of the peripheral 2H-1-benzopyran-2-one (coumarin) in metal-free and zinc(II) phthalocyanines were studied with respect to the effects on the photophysical properties of phthalocyanines. The effect of substituents on the fluorescence spectra of Pcs was also reported.

Results and discussion

8-Hydroxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin (1) and 8-hexyloxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin (2) were obtained according to the literature procedure (*16*). While metal-free (3) was obtained in the presence of 1,8-diazabicyclo-[5,4,0]-undec-7-ene (DBU)

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in anhydrous 2-(dimethylamino)-ethanol solvent, metallophthalocyanines (**4** and **5**) bearing fluorescence coumarin were accomplished by heating a mixture of 8-hexyloxy-3-[*p*- (3',4'-dicyanophenoxy)-phenyl]coumarin with anhydrous Zn(O₂CMe)₂ salts and CoCl₂ at *ca.* 180°C under N₂ atmosphere in the presence of DBU in hexanol (Scheme 1).



Scheme 1. Synthetic route of **3–5**, {M[Pc(OBzCou)₄] (M = 2H, Zn(II), Co(II))} and their lactone ring-opening complexes (**3a**, **4a**). (i) 2,3-Dihydroxybenzaldehyde, (Ac)₂O, NaOAc, 160–170°C; HCl, MeOH. (ii) C₆H₁₃Br, K₂CO₃, DMF, 80–90°C. (iii) and (iv) *N*,*N*-dimethylaminoethanol, DBU and anhydrous Zn(O₂CMe)₂, CoCl₂ and DBU. Li, *n*-hexanol, bromohexane, 24 h.

In addition, the lactone rings of 2,9,16,23-tetrakis[8hexyloxy-3-(4-oxyphenyl)coumarin]metal-free phthalocyanine (3) and 2,9,16,23-tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]phthalocyaninatozinc(II) (4) were opened and the hydroxyl groups released were hexylated. The yields of **3**–**5** were rather low (33% for **3**, 73% for **4** and 42% for **5**). The structure of phthalocyanines (3-5) was verified by FT-IR, ¹H NMR, UV-vis and MALDI-TOF MS spectroscopic methods, as well as by elemental analysis. All the analytical and spectral data are consistent with the predicted structures. Conversion of the 8-hexyloxy-3-[p-(3',4')-dicyanophenoxy)phenyl]coumarin to the phthalocyanines (3-5) was confirmed by the disappearance of the sharp $-C \equiv N$ vibration at $2230 \,\mathrm{cm}^{-1}$ in compound **2** and appearance of the strong C=O (lactone) at ca. 1701 as well as the others (Ar-H and Aliph-H together with Ar-O-Ar band) in Pcs. However, ester stretching bands in the FT-IR spectrum of 3a and 4a at *ca.* 1710 cm^{-1} are the evidence for the formation of **3a** and 4a.

The ¹H NMR spectra of compounds 3-4 are somewhat broader than the corresponding signals in the starting dinitrile derivative. This broadening is likely due to chemical exchange caused by aggregation-disaggregation equilibria and the fact that the product obtained in these reactions is a mixture of positional isomers that are expected to show chemical shifts that differ slightly from each other (11, 13). The inner NH protons of 3 and 3a were also identified in the ¹H NMR spectrum with a broad chemical shift between $\delta = -3.47$ and -3.48 ppm, respectively, as a consequence of the 18 π -electron systems of the planar molecule and this signal disappears on deuterium oxide exchange (11, 16). The other resonances related to aliphatic and aromatic (OCH₂, CH_2 , CH_3 and Ar-H) protons in the ¹H NMR spectra **3** and 3a are similar to each other except esteric protons. The ¹H NMR spectrum of **3a** and **4a** indicated characteristic aromatic (Bz₁, Bz₂ and Bz₃) at 8.40–6.78 and 8.36–6.82, ethylenic protons at 6.63 and 6.70 and aliphatic ester protons at 4.30-4.00, 4.30-4.02 ppm, respectively. The signals belonging to aromatic protons in the downfield of the spectrum in 3, 3a, 4 and 4a are broader than the aliphatic ones in the high field of the spectrum.

While divalent **4** and **4a** gave a single Q-band of high intensity due to a single $\pi - \pi^*$ transition as expected as a result of this D_{4h} symmetry, **3** and **3a** exhibited splitted Q-band as a result of D_{2h} symmetry with shoulders at slightly higher energy side of the Q-band for each phthalocyanine in addition to B-bands in UV region (17, 18). The Q-band absorptions in the DMSO of **3**, **4** and **5** were observed at 703, 675 (Qx and Qy), 683 and 669 nm, respectively, with shoulders at slightly higher energy side of the Q-band for each phthalocyanine (Figure 1). The B-bands are broad due to the superimposition of the B1- and B2-bands in the 330–350 nm region in the case of compounds **3–5** (19, 20).



Figure 1. UV-vis spectra of 3, 3a, 4, 4a and 5.

Protonated molecular ion peak for 3 (2,5-dihydroxybenzoic acid, DHB) was observed at 1858 Da for the monoisotopic masses of the elements in compound 3 (H_2Pc) with the other isotopic peaks (between 1857 and 1460 with about 1 Da mass differences). It was mainly resulted from the isotopic distribution of carbon, which was exactly overlapped with the mass of the ligand calculated theoretically from the elemental composition (13, 14). The mass spectra of compound 3a also confirmed the proposed structures giving MS (MALDI-TOF, DHB as matrix) at m/z: 2603.43 [M+H]⁺. The protonated molecular ion peaks were identified at m/z: 1922.09 $[M + H]^+$ for 4 at m/z: 2666.34 $[M + H]^+$ for 4a and at m/z: 1915.07 [M]⁺ for 5 by using DHB as matrix in MS MALDI-TOF. Following the protonated molecular ion peak, a peak group was observed at 38 Da mass higher than the protonated molecular ion peak group. This peak is resulted from the potassium adduct to the neutral molecule of 4 (Figure 2).

Fluorescence measurements

The excitation spectra of phthalocyanine derivatives with coumarin substituents were obtained at 460 and 730 nm emission wavelengths (Figure 3). The spectra at lowwavelength regions belong to coumarin substituents since the contribution of Pc is negligible (21, 22). As shown in Figure 3, the coumarin substituent exhibits high excitation intensity alone and excitation maximum wavelength at 340 nm. Maximum wavelength of coumarin moiety that bound to Pc core shifts to 1, 4, 3 and 5 nm blue region for 3, 3a, 4 and 4a, respectively. On the other hand, drastic diminishes at excitation intensities were observed in the case of 4 and 4a species. This shows that chemically treated coumarin moiety, which has a ring-opened structure, has restricted conjugation structure causing a decrease in excitation intensity (23). When monitoring the emission at 730 nm, the excitation spectra (at highwavelength regions) of coumarin-substituted 3-4 were obtained (Figure 3). It was observed that the excitation spectra of 3 and 3a closely match the absorption spectra



Figure 2. Positive ion and reflectron mode MALDI-MS spectrum of 4 (in DHB as matrix). Inset: spectrum shows expanded molecular mass region of 4.

from 450 to 730 nm. This indicates the presence of a single species and that the energy-transfer process is nearly quantitative. This also shows that **4** and **4a** are essentially in the non-aggregated form in this concentration (24). Whereas excitation spectra of coumarin-substituted **3** and **3a** are not nearly identical with their ground-state absorption spectra, showing that these Pc derivatives are in the aggregated form in this media (25, 26). Fluorescence excitation and emission spectra of compound **5** were excluded since cobalt ion in the centre of Pc almost quenched the fluorescence intensity of Pc derivative.

Figure 4 shows the fluorescence emission spectra of coumarin moiety and Pc derivatives containing coumarin substitutes in DMSO at room temperature upon excitation at 390 nm. Emissions observed at 480 nm are almost exclusively from coumarin-substituent bound to Pc core since unsubstituted Pc does not exhibit any fluorescence emission (data not shown) in this excitation wavelength. Spectrofluorimeter shows secondary harmonic emission at the wavelength of 680 nm when excitation wavelength of 340 nm used where coumarin moiety has maximum excitation intensity. Therefore, excitation wavelength was chosen as 390 nm to eliminate this emission and also to obtain emissions from both the coumarin moiety and the Pc core. As shown in Figure 4, emission intensity of coumarin

moiety is less than that of Pc derivatives because of low excitation intensity at this excitation wavelength. Whereas coumarin moiety shows higher emission intensity compared with the Pc derivatives when it was excited at 340 nm (spectra not shown). This shows that the fluorescence intensity from the peripherally bonded coumarin derivatives is decreased because of inner filter effect of Pc core causing to radiative energy transfer. The decrease



Figure 3. Excitation spectra of coumarin-substituent alone (a), 3 (b), 3a (c), 4 (d) and 4a (e) in DMSO. Inset: excitation spectra were recorded for emission at 460 and 730 nm.



Figure 4. Emission spectra of coumarin-substituent alone (a), **3** (b), **3a** (c), **4** (d) and **4a** (e) in DMSO (emission excited at 390 nm). Inset: emissions obtained for excitation at 620 nm.

in emission is more intense in the case of **3**, since **4** core gives poorer spectral overlap compared with that of metalfree core (27, 28). On the other hand, decrease in the emission intensity of coumarin derivative, which was chemically treated, is more pronounced because of the degradation of aromatic structure of coumarin moiety. The spectra of Pc derivatives containing coumarin moiety show Stokes shifts, indicating very little molecular rearrangement in the excited state. The emission maximum for coumarin moiety is blue shifted (by 15 nm) compared with coumarin-containing Pc derivatives, indicating that there are intramolecular interactions causing excited-state degeneracy as a result of molecular rearrangement (29, 30).

The fluorescence emissions at higher wavelength occur exclusively from the Pc moiety (Figure 4, right spectra). As shown in Figure 4, zinc phthalocyanine (ZnPc) species with same coumarin moieties exhibit more intense emission than those of metal-free species and there is no spectral shift in the emission wavelengths. When the steady-state fluorescence spectra of coumarin-substituted Pcs with Zn and metal-free performed in DMSO upon excitation at 620 nm Q-band vibrations, same emission spectra around 680 nm were obtained (Figure 4, inset). This shows that there is no spectral shift effect of coumarin derivatives on the fluorescence emission from S1 singlet excited state of the Pc core with or without metal ion. On the other hand, decrease in the emission of Pc core with the different type of appended coumarin derivatives suggests that the Pc core excited state rapidly decays through radiationless conversion to the ground state (31, 32).

As a conclusion, we have synthesised novel Pc derivatives possessing the axial 8-hexyloxy-3-(4-oxy-phenyl)coumarin and the cleavage of the lactone ring of coumarin substituent. These substituents enhance the solubility of Pc derivatives and inhibit the aggregation. Spectrophotometric measurements revealed that ZnPc derivatives, which have coumarin derivatives and

cleavage of its lactone ring, are in the unaggregated form, whereas H₂Pc species possessing the same substituents are in the aggregated form. This shows that not only the substitution of coumarin derivatives effects the aggregate formation, but also zinc ion in the centre of Pc prevents the cluster formation. On the other hand, derivatives of the coumarin units produced emission from them and also from Pc core upon specific excitation. It was concluded that the derivatives of coumarin units decay by two pathways: either by direct luminescence giving rise to the band at $\lambda_{max} = 485$ nm or by energy transfer to the Pc core, leading to red emission at $\lambda_{max} = 680$ nm.

Experimental

Tetrahydrofuran, chloroform (CHCl₃), Zn (O₂CCH₃)₂, CoCl₂, DBU and 4-nitro-1,2-dicyanobenzene were purchased from Merck (Darmstadt, Germany) and Alfa Aesar (Karlsruhe, Germany), and used as received. All other reagents were obtained from Aldrich Chemical Co. (Milwaukee, WI, USA) and used without purification. The purity of the products was tested in each step by thin layer chromatography. Routine IR spectra were recorded on a Shimadzu Fourier Transform FT-IR-8300 Infrared Spectrophotometer using KBr pellets, and electronic spectra on a Shimadzu UV-1601 UV-vis Spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Bruker 300 spectrometer instrument using tetramethylsilane as an internal reference at Sakarya University. Elemental analysis was performed by the Instrumental Analysis Laboratory of TUBITAK Ankara Test and Analysis Laboratory. Mass spectra were performed on a Bruker Autoflex III mass spectrometer. Fluorescence excitation and emission spectra were recorded on a Jasco FP-750 spectrofluorimeter. Emission and excitation spectra at 5 nm bandpass in both excitation and detection paths were measured in the DMSO solution containing 10 µM concentration of phthalocyanine derivatives.

8-Hydroxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin (1)

A mixture of 2,3-dihydroxybenzaldehyde (0.99 g, 7.19 mmol), p-(3,4-dicyanophenoxy)-phenylacetic acid (2.00 g, 7.19 mmol), anhydrous acetic anhydride (20 ml) and sodium acetate (2.95 g, 36 mmol) was heated and stirred at reflux temperature in a sealed glass tube for 6 h under N₂. After cooling to room temperature, water was added and the mixture stirred for one night. The resulting solid 8-acetoxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]-coumarin was filtered, washed with water and dried. The crude product 8-acetoxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin was suspended in methanol. A 10% HCl was added to make pH 3.0 and the mixture was heated

and stirred at 60°C for 4 h under N₂. At the end of the reaction, the resulting powder 8-hydroxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin was filtered, washed with water and dried. It was purified on silica gel column chromatography with CHCl₃ as an eluent.

Yield: 1.45 g (53%); mp 265–267°C. IR ν (cm⁻¹): 3387 (OH), 3074-3044 (Ar-H), 2916-2851 (alkyl-CH), 2237 (-C=N), 1709 (C=O lactone), 1616 (C=C), 1589-1485 (Ar-C=C), 1258 (Ar-O-Ar). ¹H NMR (DMSO-*d*₆) δ: 9.82 (Ar-OH, 1H, deuterium exchangeable), 8.24 (Bz₁; dd, 1H, meta to CN; H2), 8.10 (d, isomer, 1H, ortho to CN; H3), 7.88 (d, 1H, ortho to CN, phenyl H2), 7.77 (s, 1H, meta to lactone, Cou-H), 7.41 (Bz₂; dd, 4H, Ar-H), 7.23 (d, 1H, meta to Cou-OH), 7.01 (dd, 1H, ortho to Cou-OH), 6.90 (d, 1H, ortho to Cou-OH). ¹³C NMR (300 MHz, δ , DMSO- d_6): 162.3 (Ar-C=O), 161.4 (Ar-H, ortho to Ar-OH), 145.6, 142.0 (Cou), 140.8 (Bz₁), 137.9 (Bz₂), 135.5, 134.3, 133.2, 130.7, 127.3 (Bz), 127.2, 125.4 (Cou), 123.5, 123.1, 116.6 (Bz₁), 115.5, 115.0, 114.3, 40.4 (DMSO) ppm. Anal. calcd for C₂₃H₁₂N₂O₄ (380 g/mol): C, 72.63; H, 3.16; N, 7.37. Found: C, 72.50; H, 3.35; N, 7.34. UV-vis (DMF): λ_{max} (nm) (log ε): 324 (4.84), 382 (3.77 coumarin).

8-Hexyloxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin (2)

8-Hydroxy-3-[p-(3',4'-dicyanophenoxy)-phenyl]coumarin 1 (1.30 g, 3.42 mmol) and 1-bromohexane (0.57 g, 3.42 mmol) were dissolved in dry DMF (40 ml), and K₂CO₃ (0.47 g, 3.42 mmol) and tetrabutylammonium bromide (0.02 g, excess) as catalyst were added. After refluxing for 4 days under N₂, the reaction mixture was poured into ice-cold water and acidified with aqueous HCl solution. The precipitate formed was filtered, washed with water and the crude product dried. It was purified on silica gel column chromatography with CHCl₃ as an eluent.

Yield: 1.35 g (85.0%); mp 108–109°C. IR ν (cm⁻¹): 3067-3051 (Ar-H), 2939-2851 (alkyl-CH), 2230 (-C=N), 1720 (C=O lactone), 1605 (C=C), 1593-1481 (Ar-C=C), 1277 (Ar-O-Ar), 1245-1169 (Ar-O-C). ¹H NMR (DMSO- d_6) δ : 8.25 (Bz₁; dd, 1H, meta to CN; H2), 8.14 (d, isomer, 1H, ortho to CN; H3), 7.88 (d, 1H, ortho to CN, phenyl H2), 7.82 (s, 1H, meta to lactone, Cou-H), 7.45 (Bz₂; dd, 4H, Ar-H), 7.32 (d, 1H, meta to Cou-OR), 7.26 (dd, 1H, ortho to Cou-OR), 7.20 (d, 1H, ortho to Cou-OR), 3.96 (t, OCH₂-, 2H), 1.76-1.68 (m, $-OCH_2CH_2$), 1.44–1.20 (m, $CH_2CH_2CH_2CH_3$), 0.92 (t, CH₃). ¹³C NMR (300 MHz, δ , DMSO- d_6): 162.6 (Ar-C=O), 160.1 (Ar-H, ortho to Ar-OH), 144.4, 140.2 (Cou), 140.0 (Bz₁), 135.9 (Bz₂), 134.5, 133.5, 130.1, 129.7 (Bz), 127.3, 125.2 (Cou), 124.6, 123.5, 116.7 (Bz₁), 115.8, 115.4, 113.8, 67.6 (CH₂O-), 40.41 (DMSO), 28.8 (CH₂CH₂CH₃), 28.2 (OCH₂CH₂), 25.4 (OCH₂CH₂CH₂), 22.8 (CH₃CH₂), 14.8 (CH₃) ppm. Anal. calcd for C₂₉H₂₄N₂O₄ (464 g/mol): C, 75.00; H, 5.17; N, 6.03.

Found: C, 74.56; H, 5.01; N, 5.77; UV–vis (CHCl₃): λ_{max} (nm) (log ε): 326 (3.76).

2,9,16,23-Tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]metal-free phthalocyanine (3)

Compound **2** (0.10 g, 0.22 mmol) and anhydrous 2-(dimethylamino)-ethanol (1.0 ml) were refluxed with stirring for 48 h in a sealed glass tube under N_2 . After cooling to room temperature, methanol (2 ml) was added in order to precipitate the product. The green product was filtered and washed with water, MeOH, CH₃CN, EtOH, acetone, ethyl acetate and diethyl ether, and dried.

Yield: 0.033 g (33%); mp > 300°C. IR ν (cm⁻¹): 3298 (N–H), 3051–3036 (Ar-H), 2924–2866 (alkyl-CH), 1709 (C=O lactone), 1601 (C=C), 1504–1469 (Ar-C=C), 1269–1234 (Ar-O-Ar), 1169–1095 (Ar-O-C). ¹H NMR (DMSO-*d*₆) δ: 8.42–7.50 (Bz₁ vs. Bz₂; br, 28H), 7.40– 6.67 (br, 16H, Bz₁; Bz₃), 3.96 (t, OCH₂–, 8H), 1.78–1.72 (m, –OCH₂*CH*₂–, 8H), 1.44–1.22 (m, *CH*₂*CH*₂*CH*₂*CH*₃, 24H), 0.96 (t, CH₃, 12H), – 3.47 (s, br, NH, inner). Anal. calcd for C₁₁₆H₉₈N₈O₁₆ (1858 g/mol): C, 74.92; H, 5.27; N, 6.03. Found: C, 74.32; H, 5.22; N, 5.68; UV–vis (DMSO): λ_{max} (nm) (log ε): 336 (5.26), 642 (4.76), 675 (4.88), 703 (4.77). MS MALDI-TOF, DHB as matrix *m/z*: 1858.6 [M + H]⁺.

2,9,16,23-Tetrakis[(E)-hexyl-3-(2,3bis(hexyloxy)phenyl-2-(4-oxyphenyl)acrylate) phthalocyanine]metal-free phthalocyanine (3a)

To a solution of 2,9,16,23-tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]-phthalocyanine (**3**) (0.025 g, 0.0135 mmol) in dry DMF (5 ml), hexanol (7 ml) and lithium metal (0.0012 g, 0.17 mmol) were added. The mixture was refluxed for 24 h, and then it was cooled to room temperature. 1-Bromohexane (0.3 ml) was added to the reaction mixture and it was heated at 50°C for 3 days under N₂. After cooling to room temperature, the mixture was treated with 2 M HCl under ice. The precipitate was collected by filtration, washed with H₂O, MeOH, EtOH, CH₃CN, acetone, ethyl acetate and diethyl ether, and dried.

Yield: 0.005 g (15%); mp > 300°C. IR ν (cm⁻¹): 3290 (N–H), 3067–3044 (Ar-H), 2932–2866 (alkyl-CH), 1720 (C=O ester), 1597 (C=C), 1504–1474 (Ar-C=C), 1269–1234 (Ar-O-Ar), 1165–1096 (Ar-O-C). ¹H NMR (DMSO-*d*₆) δ: 8.40–7.22 (Bz₁ vs. Bz₂; br, 28H), 7.50– 6.78 (br, 12H, Bz₁; Bz₃), 6.63 (s, 4H, –C=CH–), 4.30– 4.00 (t, OCOC*H*₂, 8H), 3.96–3.70 (t, OC*H*₂–, 16H), 1.80–1.60 (m, –OCH₂C*H*₂–, 24H), 1.43–1.24 (m, *CH*₂*CH*₂*CH*₂CH₃, 72H), 0.94 (t, CH₃, 36H), –3.48 (s, br, NH, inner, 2H) ppm. Anal. calcd for C₁₆₄H₂₀₂N₈O₂₀ (2602 g/mol): C, 75.63; H, 7.76; N, 4.30. Found: C, 74.77; H, 7.28; N, 4.18; UV–vis (DMSO): λ_{max} (nm) (log ε): 291 (4.78), 330 (4.79), 639 (sh, 4.21), 680 (4.33), 704 (4.15). MS (MALDI-TOF, DHB as matrix) m/z: 2603.43 $[M + H]^+$.

2,9,16,23-Tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]phthalocyaninatozinc(II) (4)

A mixture of compound **2** (0.10 g, 0.215 mmol), DBU $(0.07 \text{ cm}^3, 0.04 \text{ mmol})$ and Zn $(\text{AcO})_2$ (0.012 g, 0.055 mmol) in hexanol (1.0 cm^3) was heated and stirred at 170°C for 18 h under N₂. The resulting green mixture was cooled to room temperature and then methanol (3 ml) was added in order to precipitate the product. The product was filtered and washed with water, methanol, acetonitrile, ethanol, acetone, ethyl acetate and diethyl ether, and dried.

Yield: 0.076 g (73%); mp > 300°C. IR ν (cm⁻¹): 3067–3044 (Ar-H), 2924–2843 (alkyl-CH), 1720 (C=O lactone), 1601 (C=C), 1500–1358 (Ar-C=C), 1269– 1234 (Ar-O-Ar), 1173–1096 (Ar-O-C). ¹H NMR (DMSO-*d*₆) δ : 8.33–7.24 (Bz₁ vs. Bz₂; br, 28H), 7.30– 6.62 (br, 16H, Bz₁; Bz₃), 3.95 (t, OCH₂–, 8H), 1.74–1.68 (m, $-OCH_2CH_2$ –, 8H), 1.45–1.25 (m, $CH_2CH_2CH_2CH_3$, 24H), 0.96 (t, CH₃, 12H). Anal. calcd for C₁₁₆H₉₆N₈O₁₆Zn (1921 g/mol): C, 72.46; H, 4.99; N, 5.83. Found: C, 72.22; H, 5.29; N, 5.55; UV–vis (DMSO): λ_{max} (nm) (log ε): 337 (4.66), 621 (sh, 3.96), 683 (4.56). MS (MALDI-TOF; DHB as matrix) *m/z*: 1922.09 [M+H]⁺, 1960.09 [M+K]⁺.

2,9,16,23-Tetrakis[(E)-hexyl-3-(2,3-bis(hexyloxy)phenyl-2-(4-oxyphenyl)acrylate)]-phthalocyaninatozinc(II) (4a)

To a solution of 2,9,16,23-tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]phthalocyaninatozinc(II) (4) (0.030 g, 0.0135 mmol) in dry DMF (5 ml), hexanol (5 ml) and lithium metal (0.0012 g, 0.171 mmol) were added. The mixture was refluxed for 24 h, and then it was cooled to room temperature. 1-Bromohexane (0.3 ml) was added to the reaction mixture and it was heated at 50°C for 3 days under N₂. After cooling to room temperature, the mixture was treated with 2 M HCl under ice. The precipitate was collected by filtration, washed with water, methanol, acetonitrile, ethanol, acetone, ethyl acetate and diethyl ether, and dried.

Yield: 0.005 g (12%); mp > 300°C. IR ν (cm⁻¹): 3036–3059 (Ar-H), 2932–2858 (alkyl-CH), 1720 (C=O ester), 1601 (C=C), 1504–1466 (Ar-C=C), 1269–1234 (Ar-O-Ar), 1161–1096 (Ar-O-C). ¹H NMR (DMSO-*d*₆) δ : 8.36–7.69 (Bz₁ vs. Bz₂; br, 28H), 7.60–6.82 (br, 12H, Bz₁; Bz₃), 6.70 (s, 4H, -C=CH-), 4.30–4.02 (t, OCOCH₂, 8H), 3.92 (t, OCH₂-, 16H), 1.84–1.53 (m, superimposed, br, -OCH₂CH₂-, 24H), 1.44–1.20 (m, br, *CH*₂*CH*₂*CH*₂CH₃, 72H), 0.94 (t, CH₃, 36H) ppm. Anal. calcd for C₁₆₄H₂₀₀N₈O₂₀Zn (2665 g/mol): C, 73.85; H, 7.51; N, 4.20. Found: C, 73.43; H, 7.41; N, 4.05; UV–vis (DMSO): λ_{max} (nm) (log ε): 349 (4.80), 621 (sh, 4.27), 660 (sh, 4.59), 682 (4.90). MS (MALDI-TOF, DHB as matrix) *m/z*: 2666.34 [M + H]⁺.

2,9,16,23-Tetrakis[8-hexyloxy-3-(4-oxyphenyl)coumarin]phthalocyaninatocobalt(II) (5)

A mixture of compound **2** (0.10 g, 0.215 mmol), DBU (0.07 ml, 0.04 mmol) and CoCl₂ (0.0134 g, 0.054 mmol) in hexanol (1.5 ml) was heated and stirred at 170° C for 24 h under N₂. The resulting green mixture was cooled to room temperature and then methanol (2 ml) was added in order to precipitate the product. The product was filtered and washed with water, methanol, acetonitrile, ethanol, acetone, ethyl acetate and diethyl ether, and dried.

Yield: 0.043 g (42%); mp > 300°C. IR ν (cm⁻¹): 3067–3036 (Ar-H), 2916–2866 (alkyl-CH), 1720 (C=O lactone), 1601 (C=C), 1504–1469 (Ar-C=C), 1323– 1234 (Ar-O-Ar), 1169–1096 (Ar-O-C). Anal. calcd for C₁₁₆H₉₆N₈O₁₆Co (1915 g/mol): C, 72.69; H, 5.01; N, 5.85. Found: C, 72.22; H, 5.14; N, 5.38; UV–vis (DMSO): λ_{max} (nm) (log ε): 335 (4.47), 606 (3,77), 669 (4.23). MS (MALDI-TOF, DHB as matrix) *m/z*: 1915.07 [M]⁺.

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